



Radiolabeling and imaging approaches for carbon-based nanohybrids

Antonis Skliris

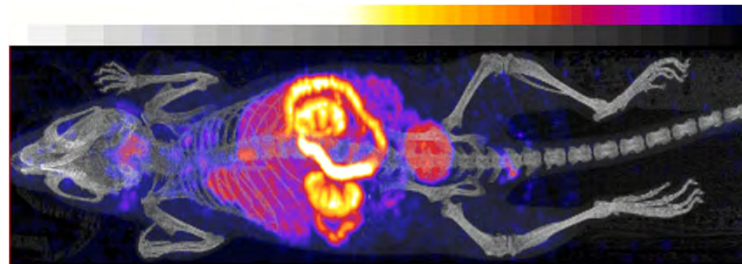
Nanohybrides 18 – Bastia May-June 2022

Who we are ?

BIOEMTECH is a SME who develops and offers innovative solutions in medical and pharmaceutical research in the **non-invasive *in vivo* imaging**.

We focus on **molecular imaging** and **biomedical engineering**:

- ✓ Design and construction of low-cost benchtop imaging devices
- ✓ Performance of preclinical imaging services in our imaging platform
- ✓ Computational simulations using Monte Carlo techniques



- R&D labs at the Technology Park "**Lefkippos**" in **NCSR Demokritos** since 2017.



In-vitro Lab

In vitro testing
(targeting,
cytotoxicity, etc.)



Radiochemistry Lab

Radiolabeling of
compounds & QC



Animal Hosting

Mouse models
(oncology, etc.)



In-vivo Imaging Lab

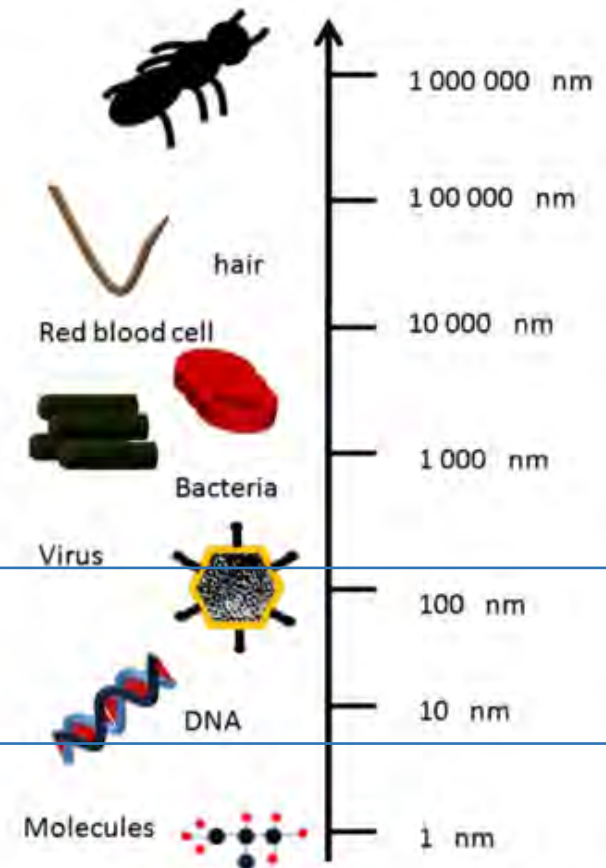
2D/3D, static or
dynamic
PET/SPECT/CT
imaging

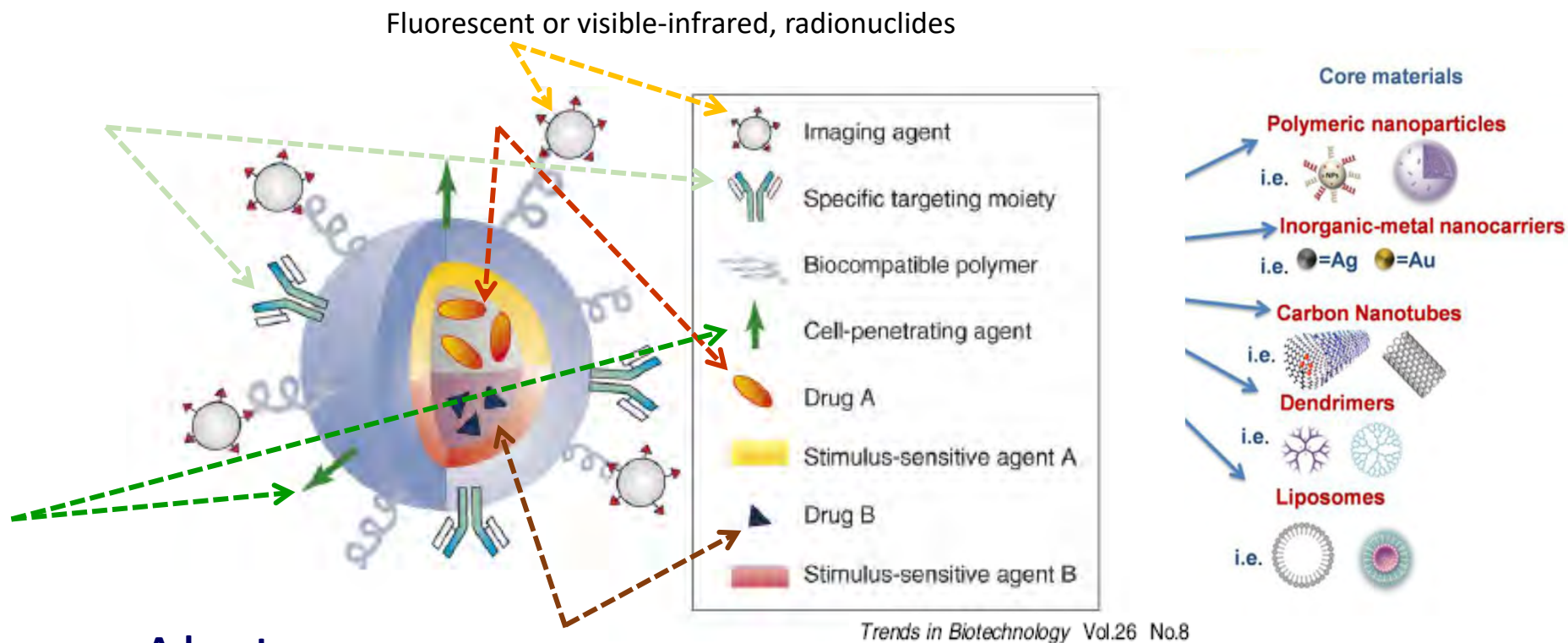
Nanoparticles

Structures where AT LEAST one dimension is between 1 and 100 nm (sometimes may be bigger)

Nanoscale

- Human: 1,800,000,000 nm
- Insect: 1,000,000 nm
- Hair: ~ 10,000 nm
- Blood cell: 8,000 nm
- Bacteria: eColi 2,000 nm
- Virus: 100 nm
- Protein: ~1-20 nm
- Molecule: ~1nm
- Hydrogen atom: 0.04 nm





Advantages:

- Selected tumor/organ targeting;
- Low drug concentration in normal tissues
- Controlled drug release in target tumor/organ
- Minimization of side effects due to lower dose and targeted delivery

BIOEMTECH | Labs



In-vitro Lab



In-vivo Imaging Lab



Radiochemistry Lab



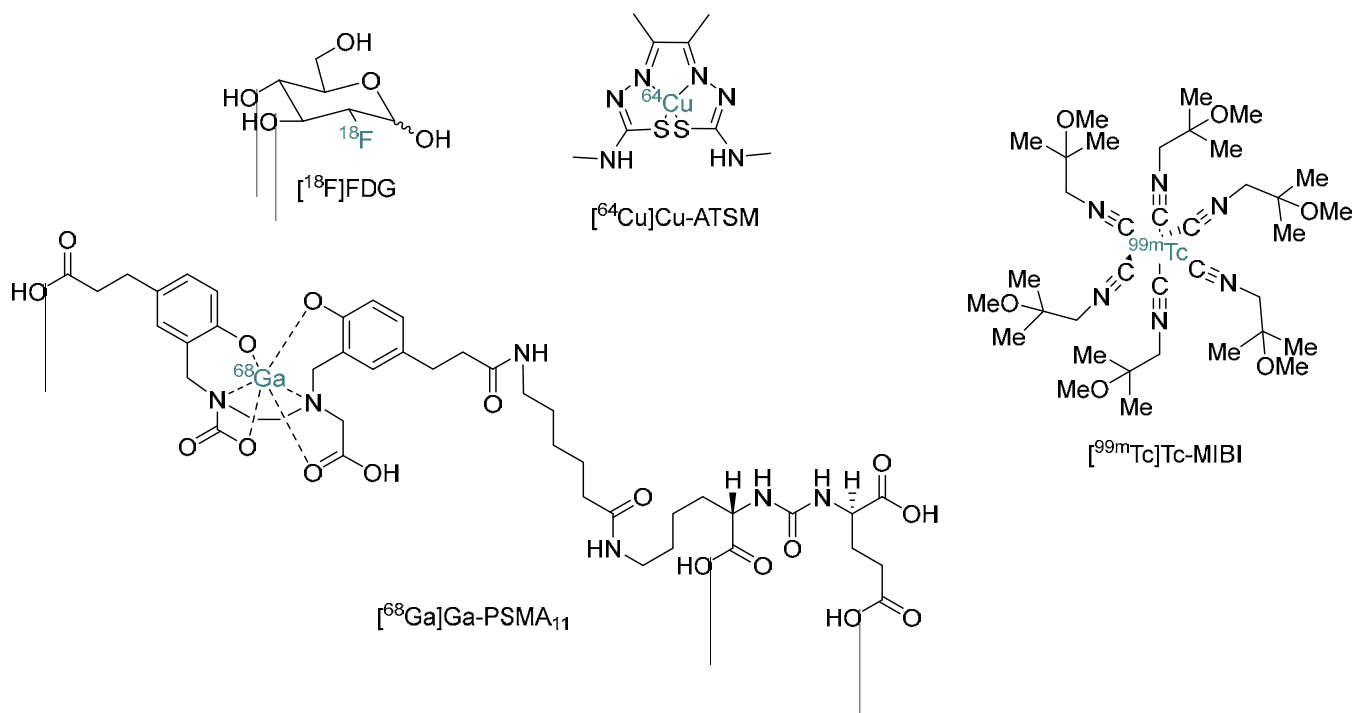
Animal Hosting

Radiolabeling of
compounds & QC

Labelling approaches

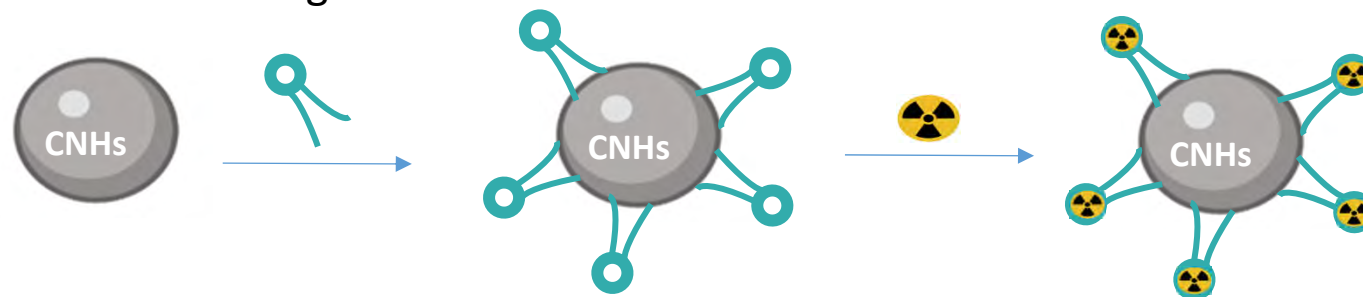
Metallic Isotopes: Use of precursors that chelate the radiometal, transmetallation reactions

Non-metallic isotopes: Use of organic chemistry reactions (SN , SN_2 , SN_{Ar} , click reactions)

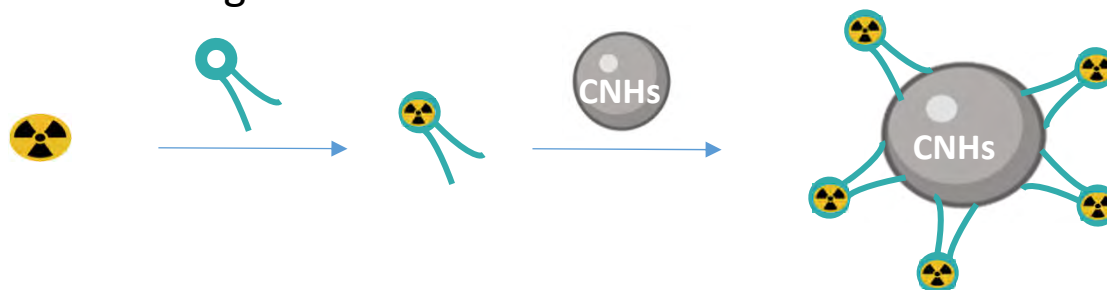


Radiolabelling approaches

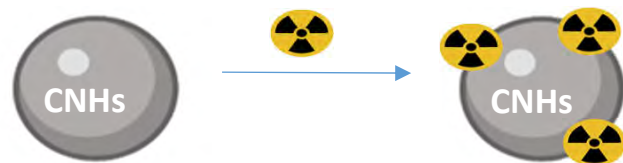
Post-radiolabeling



Pre-radiolabeling



Direct radiolabeling (no chelator)



Perform QC
Calculation of the
radiochemical conversion

Stockhofe et al., Pharmaceuticals 2014;7, 392-418

Types of radioisotopes

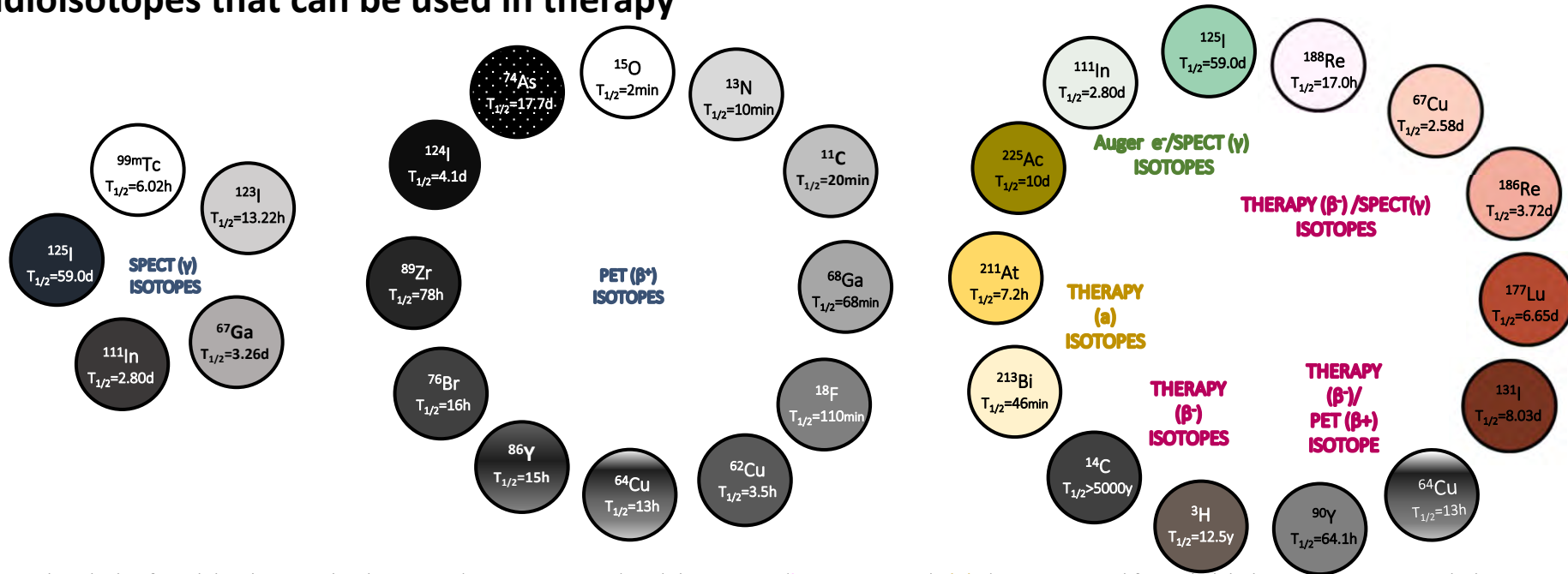
Long lived isotopes and Short lived isotopes

In medicine we usually use short-lived isotopes

Metallic radioisotopes and non metallic radioisotopes

ex. Metallic: technetium-99m gallium-68, copper-64 and zirconium-89; Non-metallic: carbon-11, fluorine-18

Radioisotopes that can be used in therapy



The Clock-Of-Nuclides showing the diagnostic (gamma, positron) and therapeutic (beta, auger and alpha) emitters used for radiolabeling NCs. At noon with the shortest physical half-life and ending with the longest physical half-life.

Analytical techniques

Radio-TLC



Radio-HPLC



γ -counter



*Measures: the percentage of the incorporated radioisotope.
It is used: synthesis, stability tests*

*Measures: the activity per sample
It is used: metabolite analysis, tracer kinetic studies *in vitro*, and pre-clinical biodistribution studies*



Scintillator detectors

Why?

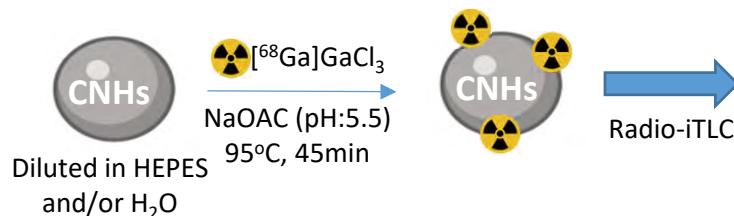
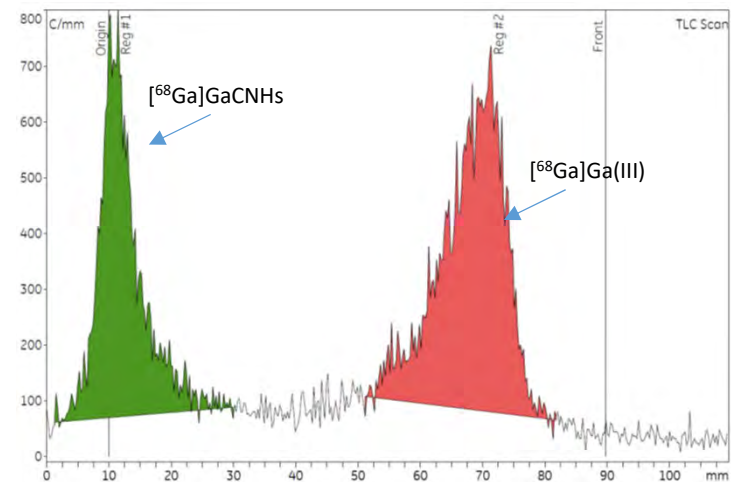
to ensure necessary chemical functionalization of the NHs for their successful in-vivo introduction

- Radiolabeling of different carbon nanohybrids (CNHs) received from the partners with $[^{68}\text{Ga}]\text{Ga}(\text{III})$ for PET imaging.

Nanomaterials tested	
CFO	carbon fluorooxide
CD3011	carbon nanodots
CDF19	carbon nanodots
S2	carbon nanodots

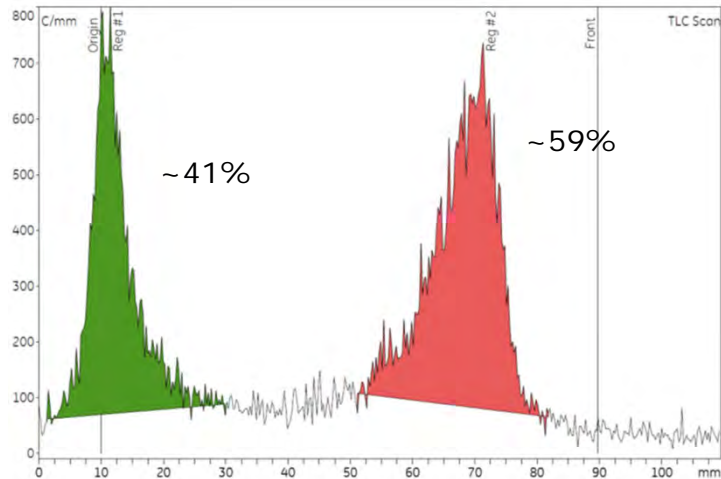


Whatman 3 MM, 0.1M EDTA

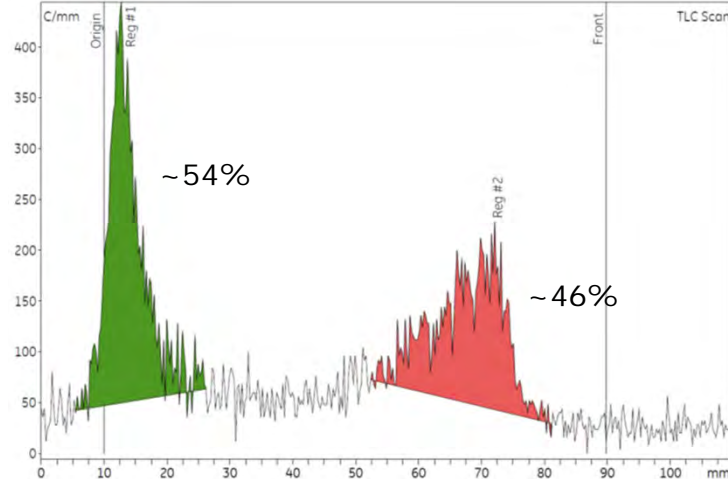


QC of radiolabeled CNHs

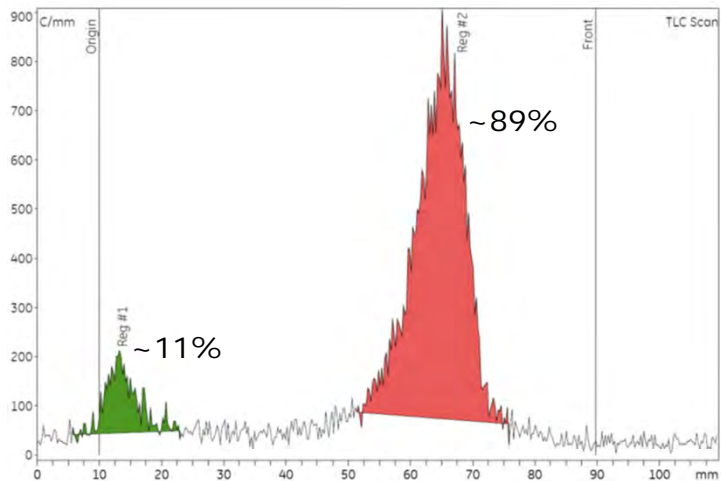
Radiochemical conversion (RCC)



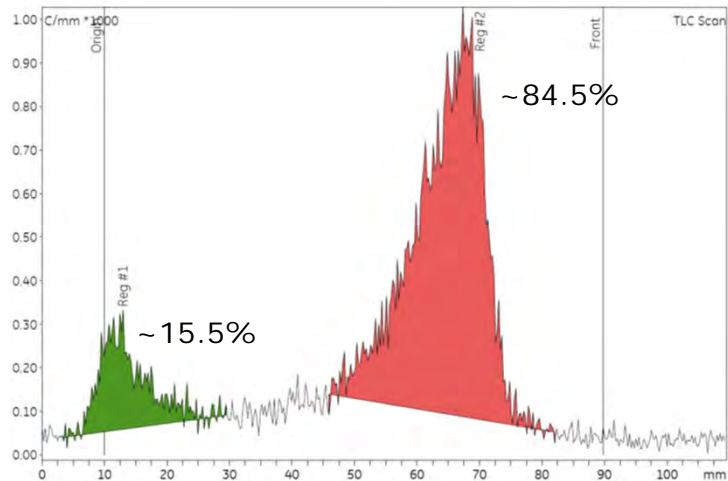
0.05mg S2 (H₂O)



0.1mg S2 (H₂O)



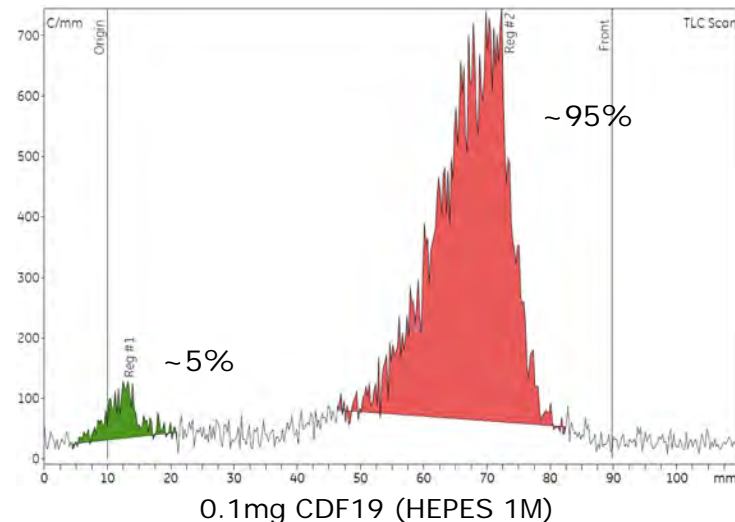
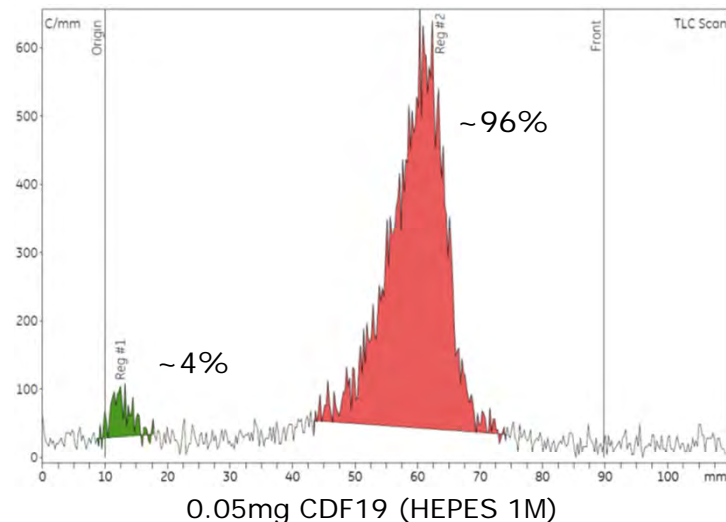
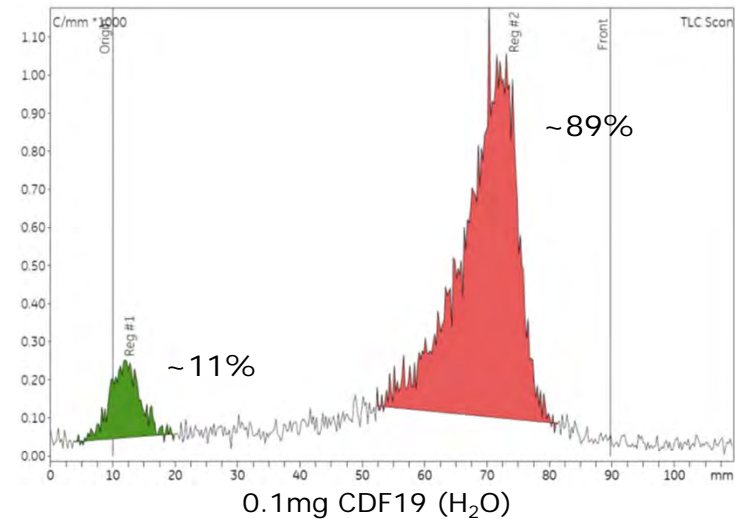
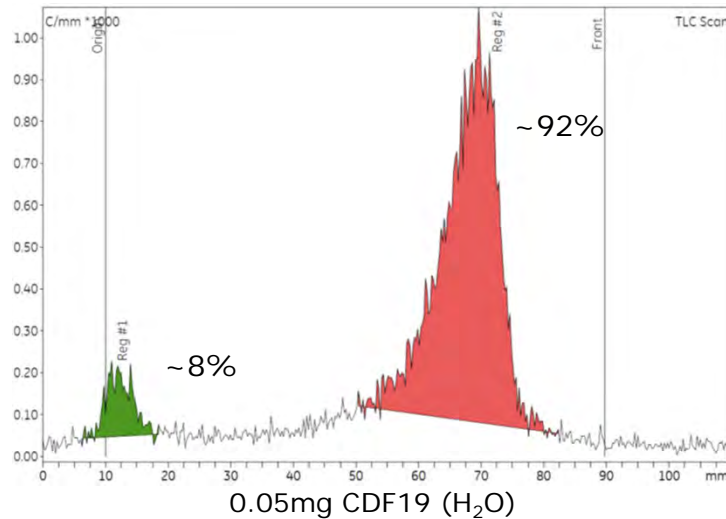
0.05mg CFO (HEPES 1M)



0.1mg CFO (HEPES 1M)

QC of radiolabeled CNHs

Radiochemical conversion (RCC)



QC of radiolabeled CNHs

Radiochemical conversion

Findings

- Radiochemical conversion up to ~54% (for S2)
- Low radiochemical conversion for the other CNHs (~3% - 15.5%)
- CNH dilution buffer & amount of CNHs affect radiolabeling



What is needed

- Optimization of the $[^{68}\text{Ga}]\text{GaCl}_3$ label protocol (pH, buffer ect.)
- Label nanomaterials with $[^{111}\text{In}]\text{InCl}_3$ for SPECT imaging



Next step

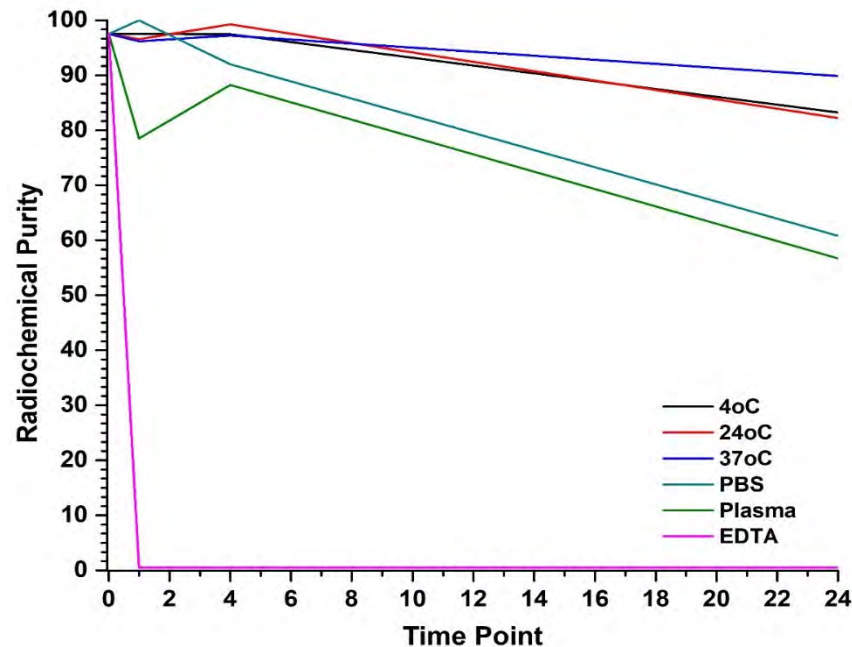
- Perform kinetics stability (temperatures, buffers)

QC of radiolabeled CNHs

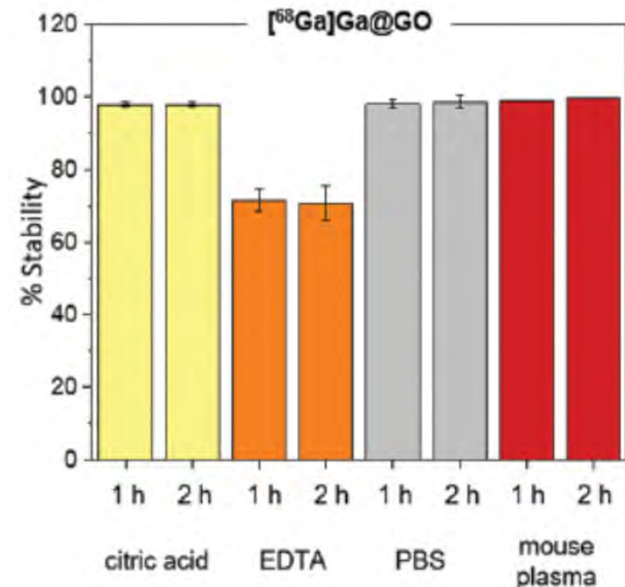
Kinetic Stability Assays

Aliquots of the reaction mixture were taken and incubated at different temperatures and different solvents

Radio-TLCs were taken at 1hr, 4 hrs and 24hrs post preparation.



Comparative diagram probing the potential *in vivo* stability of the radiolabeled-NHs at three time points post-preparation (p.p.) under various incubation conditions (PBS, EDTA, Plasma).



What's next

If:

- $RCC \leq 95\%$
- Low stability over time



Then:

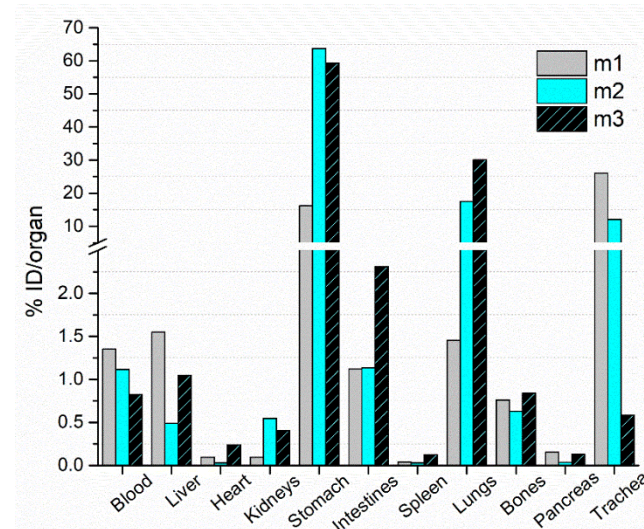
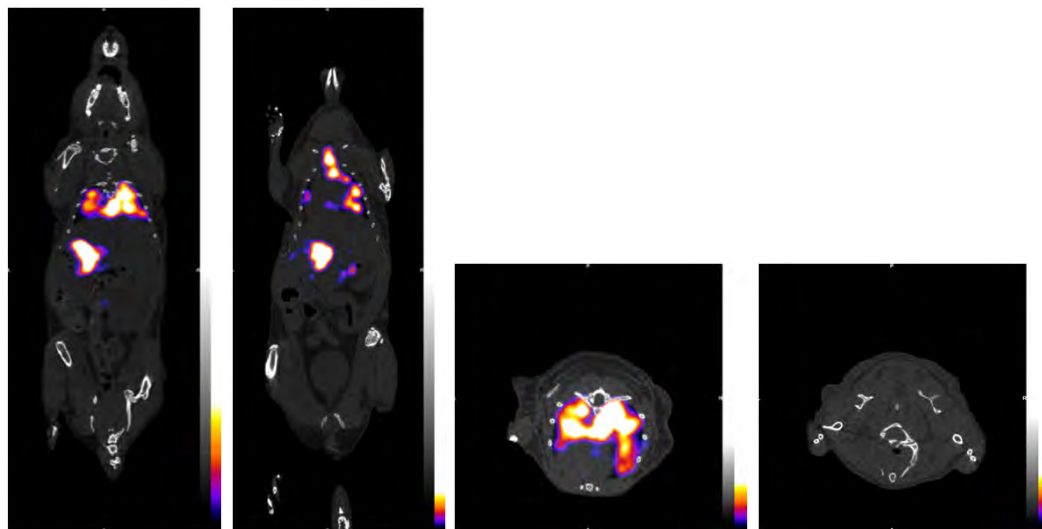
Return to nanomaterial preparation to improve characteristics and performance

- $RCC \geq 95\%$
- Stable over time



In vivo administration

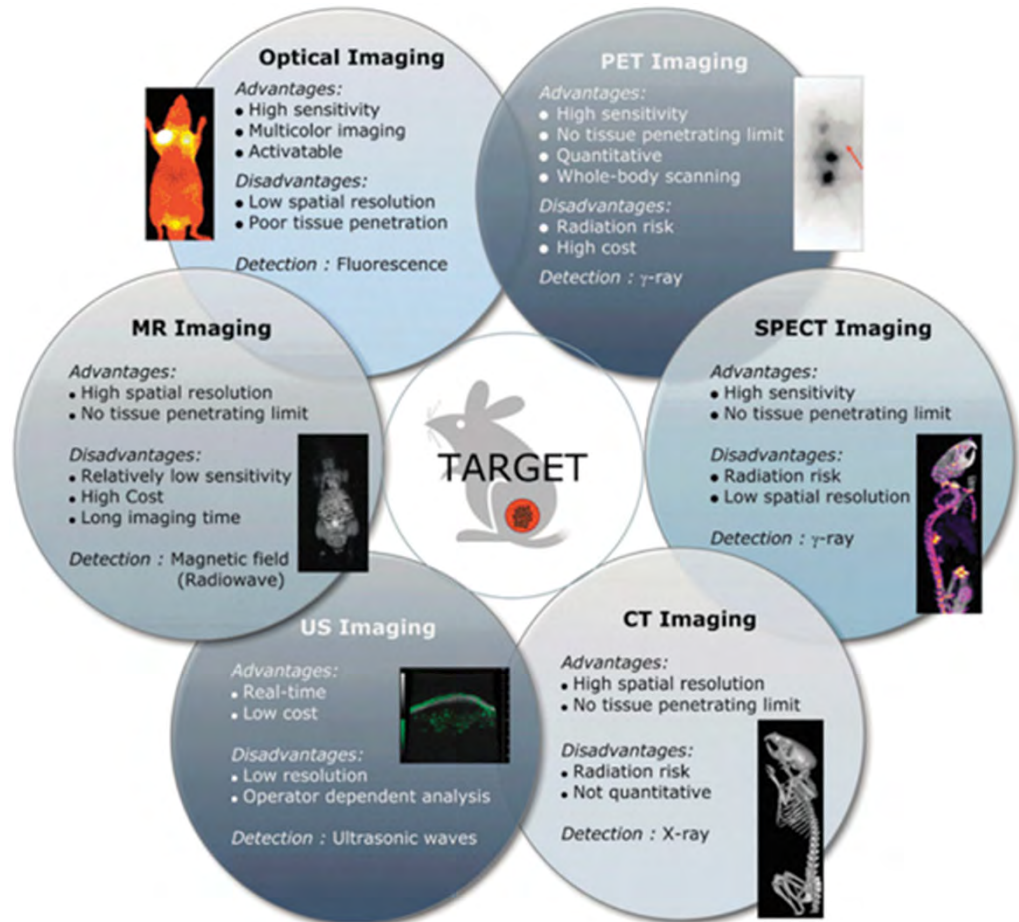
- Imaging studies
- Biodistribution studies



Our Imaging Platform

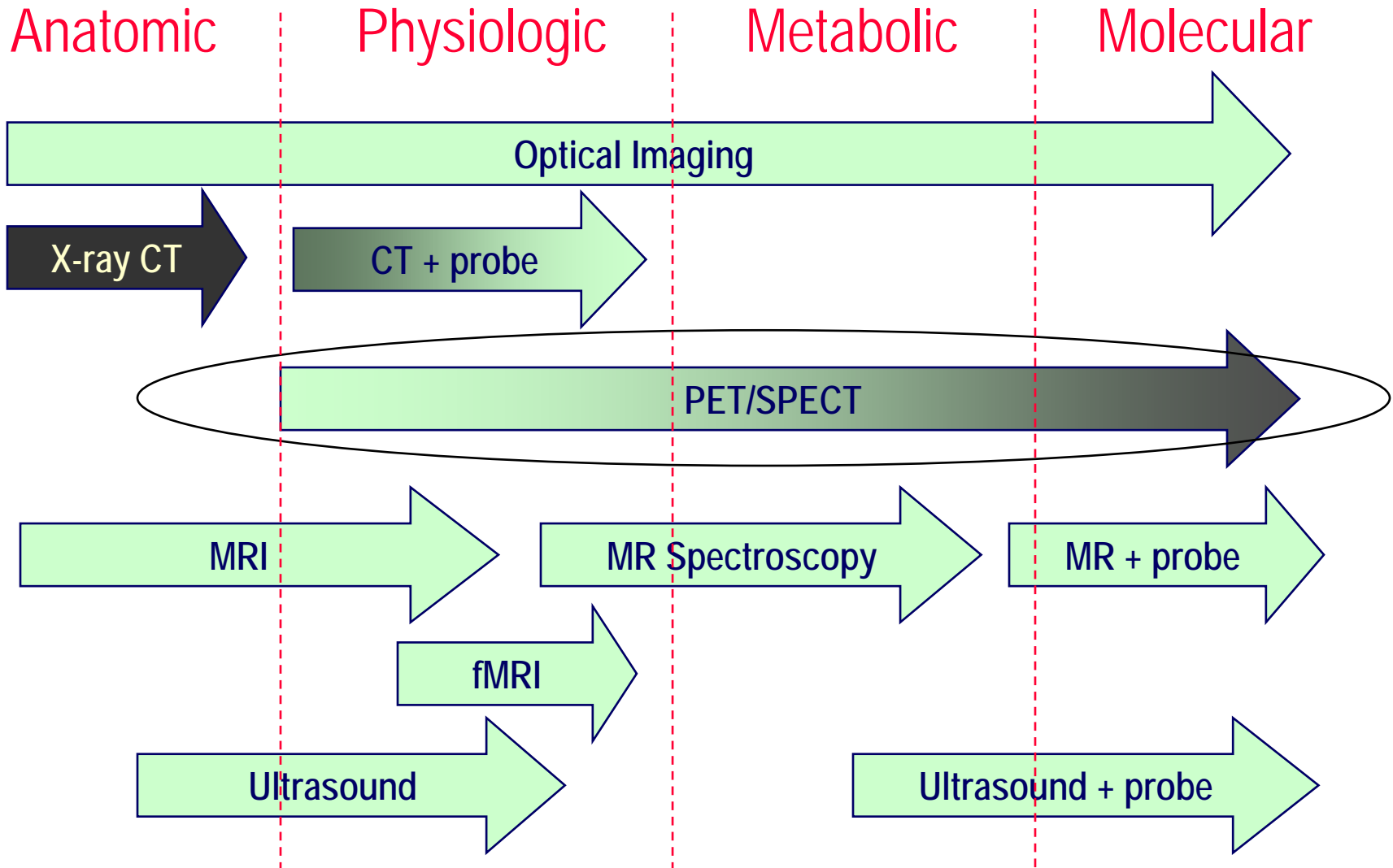
Small animal imaging

- In vivo \neq In vitro
- Non-destructive. Repeated studies in the same animal
- Each animal serves as its own control
- We can efficiently image the entire animal simultaneously
- Imaging bridges the gap from cell to human studies
- Many potential targets
- A large variety of imaging techniques available

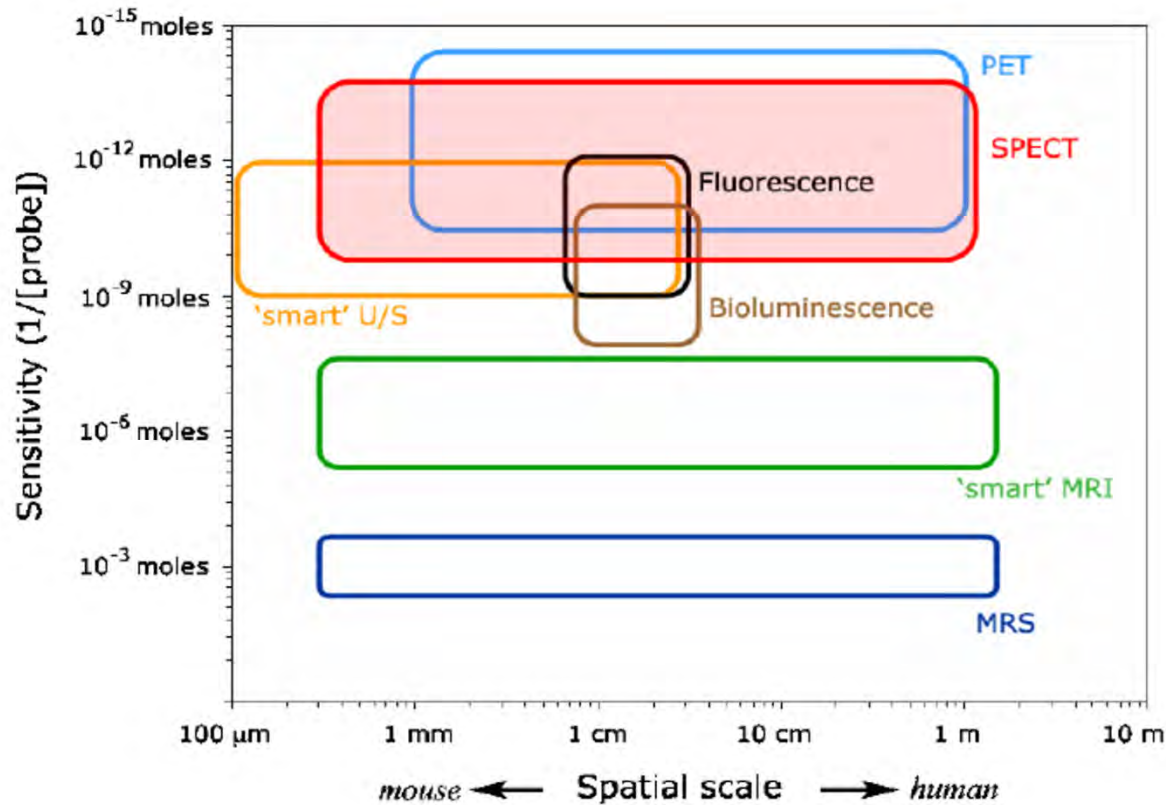


A Molecular Imaging Primer: Modalities, Imaging Agents, and Applications, Michelle L. James and Sanjiv S. Gambhir, Physiological Reviews, Vol. 92, No. 2, 01 Apr 2012.

Imaging modalities



Imaging modalities



SPECT is among the most sensitive of the molecular *in vivo* imaging technologies and its spatial scale spans the resolution required for imaging small laboratory animals and the depth penetration required for imaging humans.

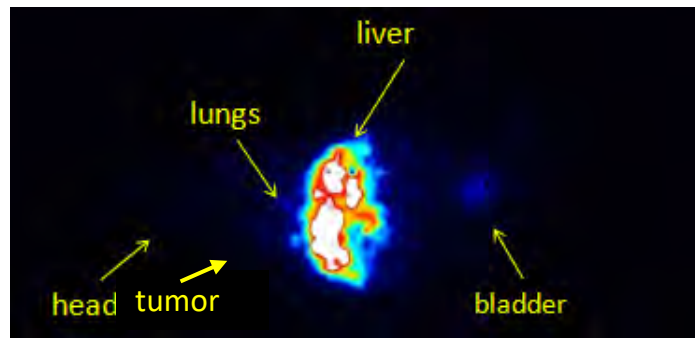
Comparison of imaging technologies

Technique	Resolution	Sensitivity	Depth	Time
MRI	10-100 μ m	μ -mMol	No limit	Min
CT	50 μ m	m-cMol	No limit	Sec
US	<50 μ m	mMol	mm	Sec
PET	1-2mm	p-nMol	No limit	Min
SPET	< 1mm	p-nMol	No limit	Min
FRI	1-2mm	p-nMol	< 1cm	Sec
FMT	1-2mm	p-nMol	< 10cm	Sec

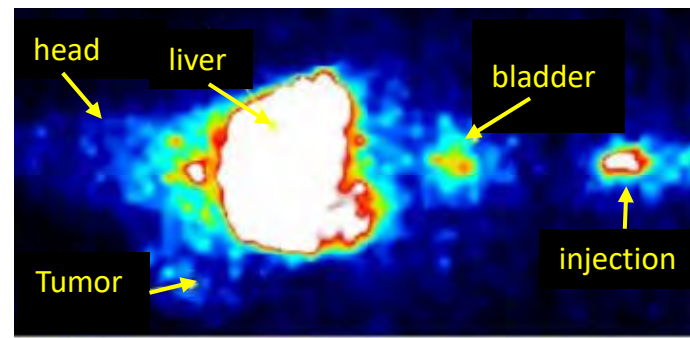
In-vivo imaging provides an answer to the following questions:

- ✓ Do nanoparticles reach the target?
- ✓ Are they concentrated in other organs/tissues?
- ✓ How long do they remain on the target?
- ✓ How long do they stay in blood circulation?
- ✓ Are they stable post injection?
- ✓ What happens at the first minutes post injection?
- ✓ What is the best injected concentration?
- ✓ How shall we prepare the animals?
- ✓ When is the highest concentration in target?
- ✓ When is the best time point for tomographic imaging?
- ✓ When is the best time for biodistribution points?
- ✓ Which is the best injection route?

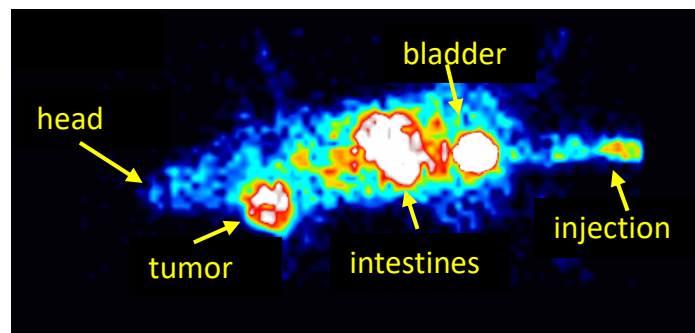
Assesment of **tumor targeting** does not need **tomographic imaging** for preliminary assessment



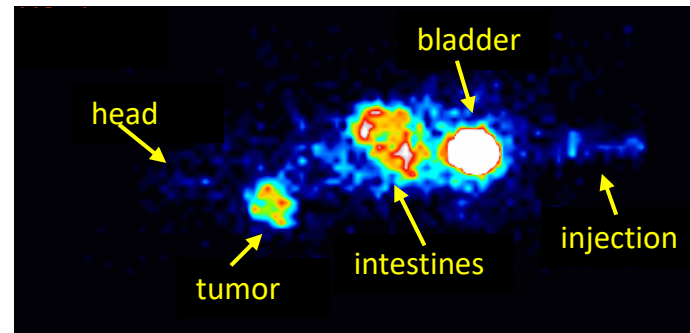
No concentration on tumor



Low concentration on tumor



Good concentration on tumor



High concentration on tumor

Different administration routes

Retro-orbital



Intra-rectal

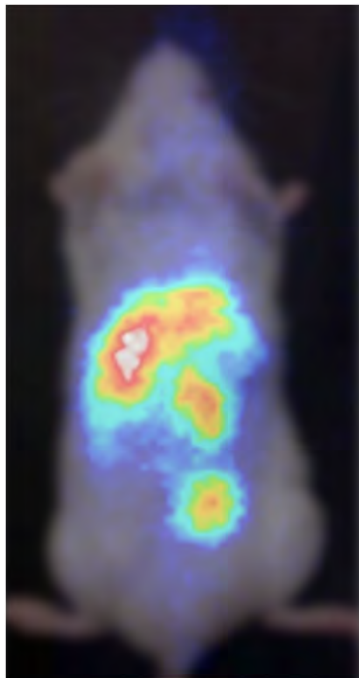


Intra-tracheal

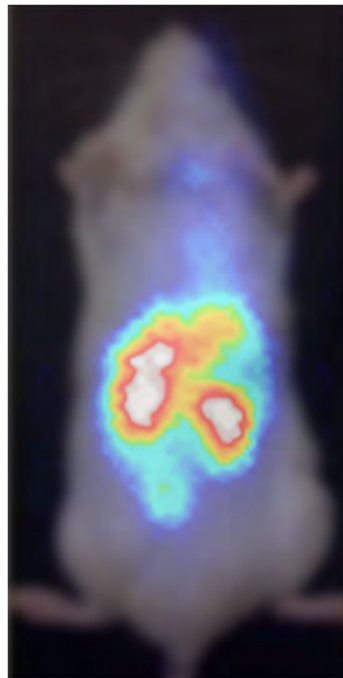


Optimize protocol parameters

Different concentrations

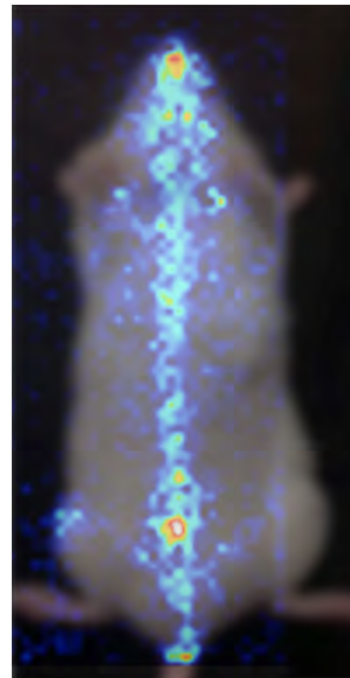


[^{99m}Tc]Tc-MIBI
100uCi
almost no heart
signal

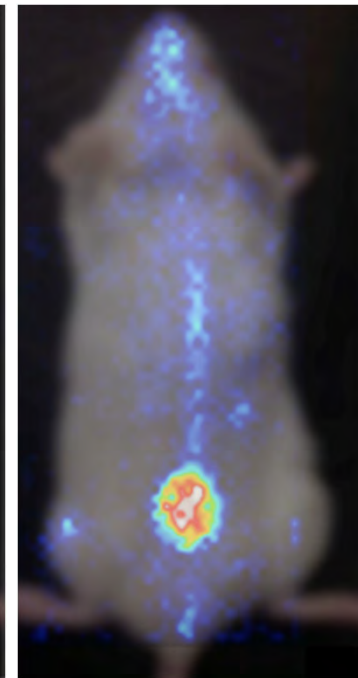


[^{99m}Tc]Tc-MIBI 2mCi
noticeable heart
signal

Different preparation conditions



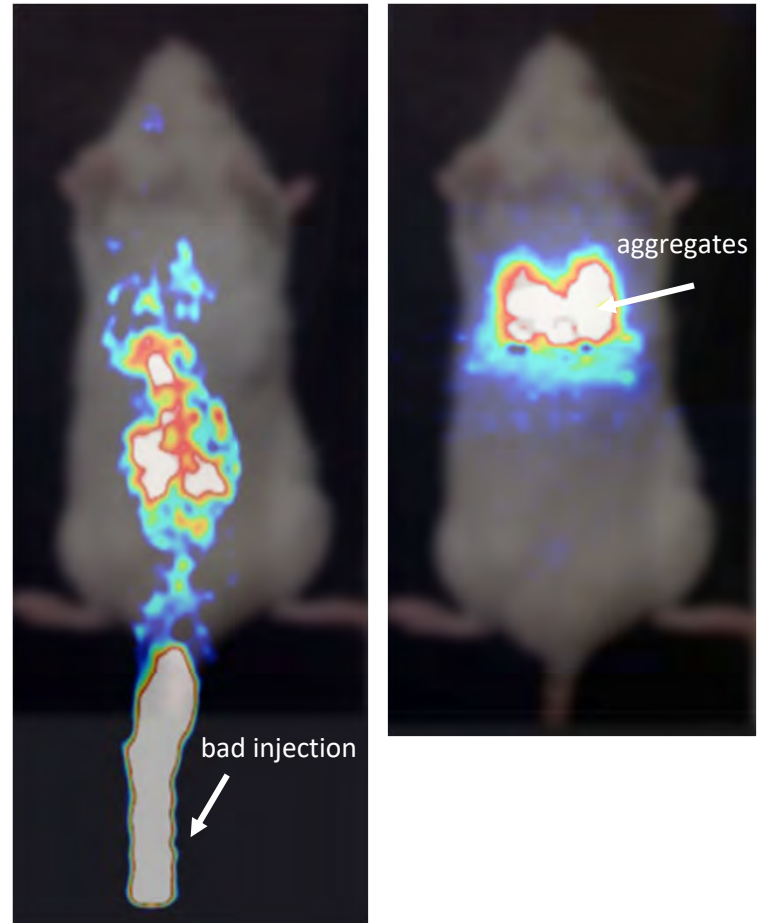
[^{99m}Tc]Tc-MDP after
water fasting
Bones clearly visible



[^{99m}Tc]Tc-MDP under
normal feeding
conditions – need to
“burn” image to see the
bones

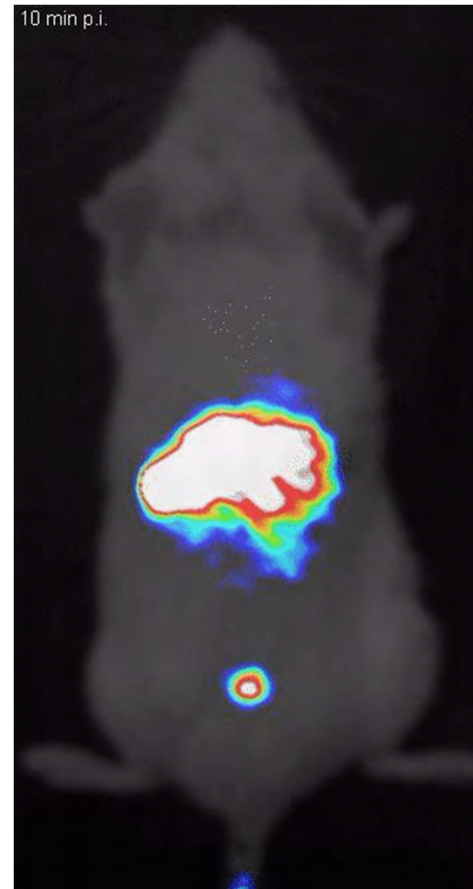
Identify unsuccessful tests

- Image all mice that will participate in a biodistribution study before dissection
- Bad injections can be identified and excluded to improve statistics
- Aggregations or other unexpected concentrations are visible
- **Animals with obvious “errors” are excluded from biodistributions**



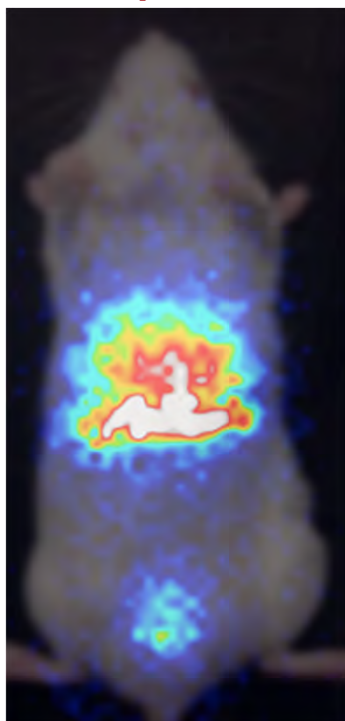
Reduction of animals

Scan the same mouse easily and fast over multiple time points, with frames down to few seconds. **Full bio distribution data with one animal**



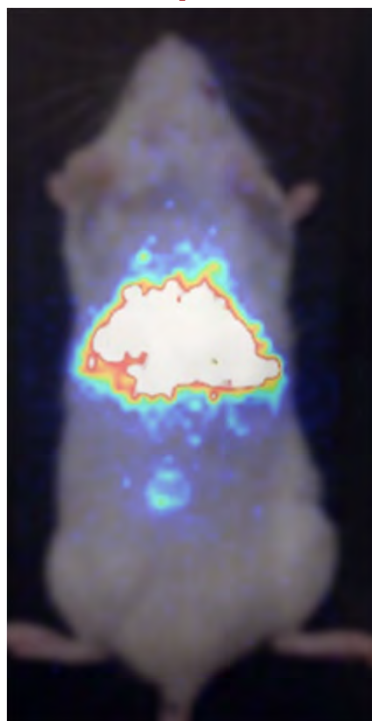
Nanoparticles Biodistribution

**Magnetic
Nanoparticles**



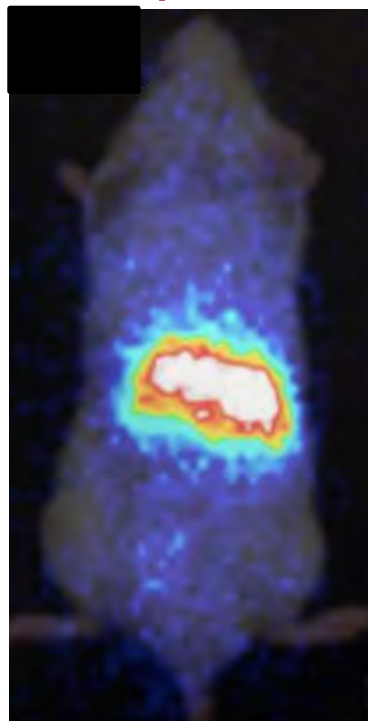
100uCi of [^{99m}Tc]Tc-MDP
in 100ul - 400 μg FeCaP
20 min acquisition

**Selenium
Nanoparticles**



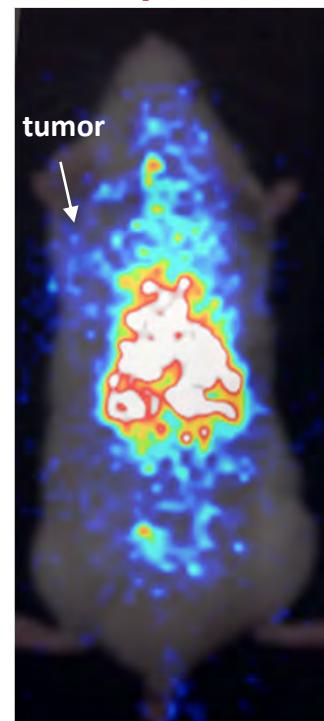
[^{99m}Tc]Tc-PLGA/SeNp
100 ul with 100 uCi
20 min acquisition

**Gold
Nanoparticles**



[^{68}Ga] Gold nanoparticles
100 ul with 10 uCi
10 min acquisition

**Liposome
Nanoparticles**



U-87MG mouse, 100 μCi
of [^{99m}Tc]Tc-NT Lipo-Cys
20 min acquisition

**Silver
Nanoparticles**



U87MG mouse, Ag-
 ^{99m}Tc (100 μCi)
20 min acquisition

Imaging examples of nanoparticles

Nanoparticles

- i. Dynamic imaging of silver NPs
- ii. SPECT imaging of magnetic NPs
- iii. CT imaging of gold NPs



Indicative preclinical studies



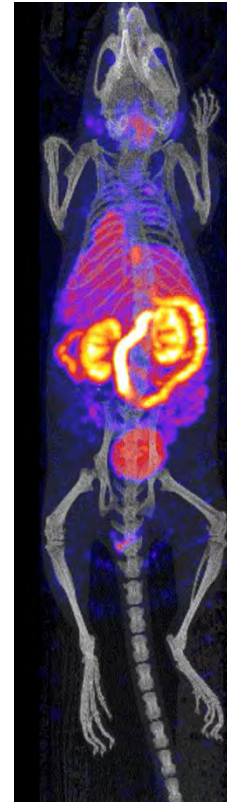
**Tumor
imaging**



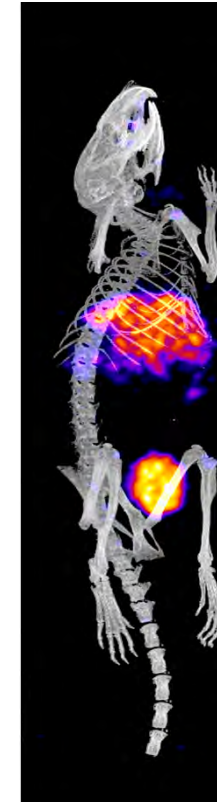
**Contrast
agents**



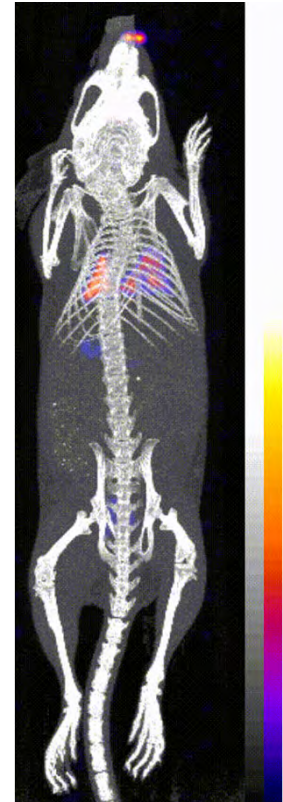
**Bone
imaging**



**Cardiac
imaging**



**Nanoparticle
imaging**

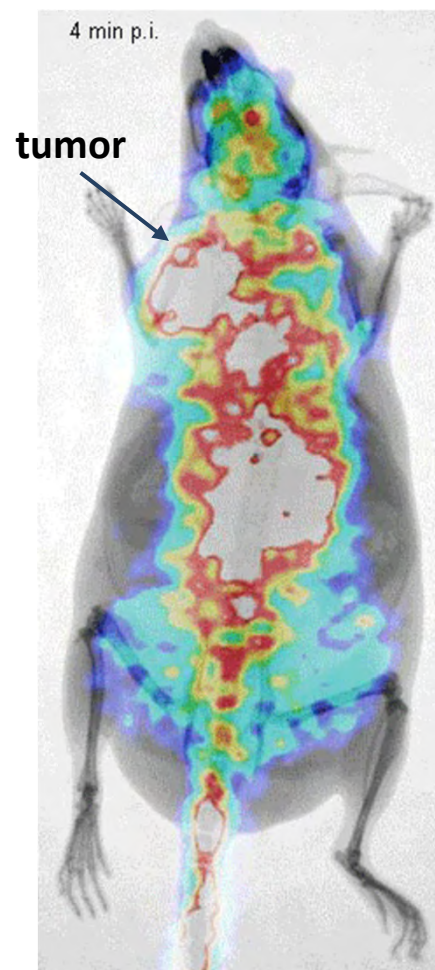


**Lung
imaging**

Tumor imaging

Tumor bearing mouse i.v. injected with 100 μ L. 1mCi [^{99m}Tc]Tc-peptide:

- i. Dynamic imaging for the first 2 hrs p.i.
- ii. 3D SPECT imaging @ 4 hrs p.i.



Imaging examples (II): Lungs

Lung imaging

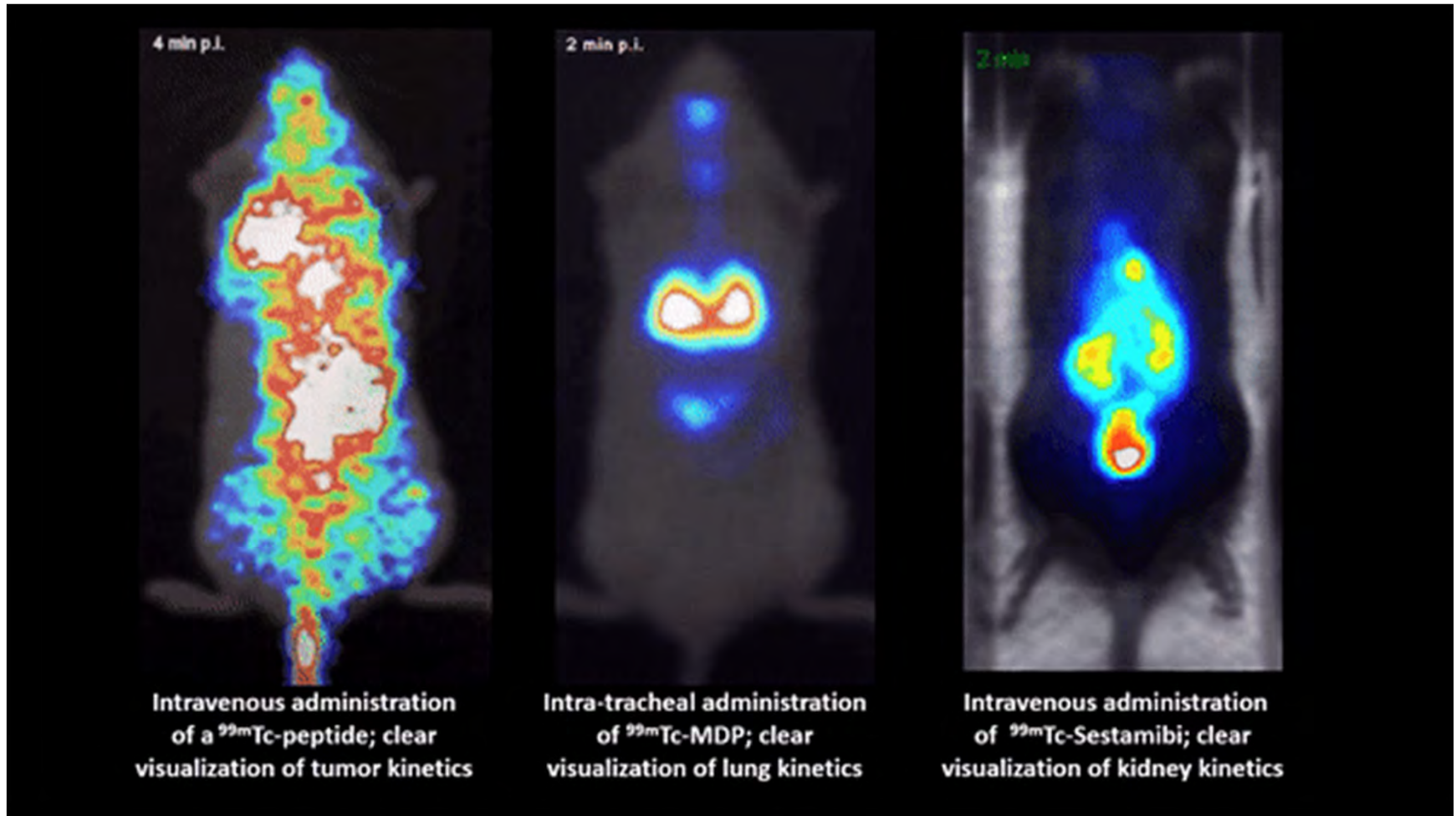
Normal mouse and intra-tracheal administration:

a) Dynamic imaging: 1 hr p.i.

b) Imaging @ 1 hrs p.i.

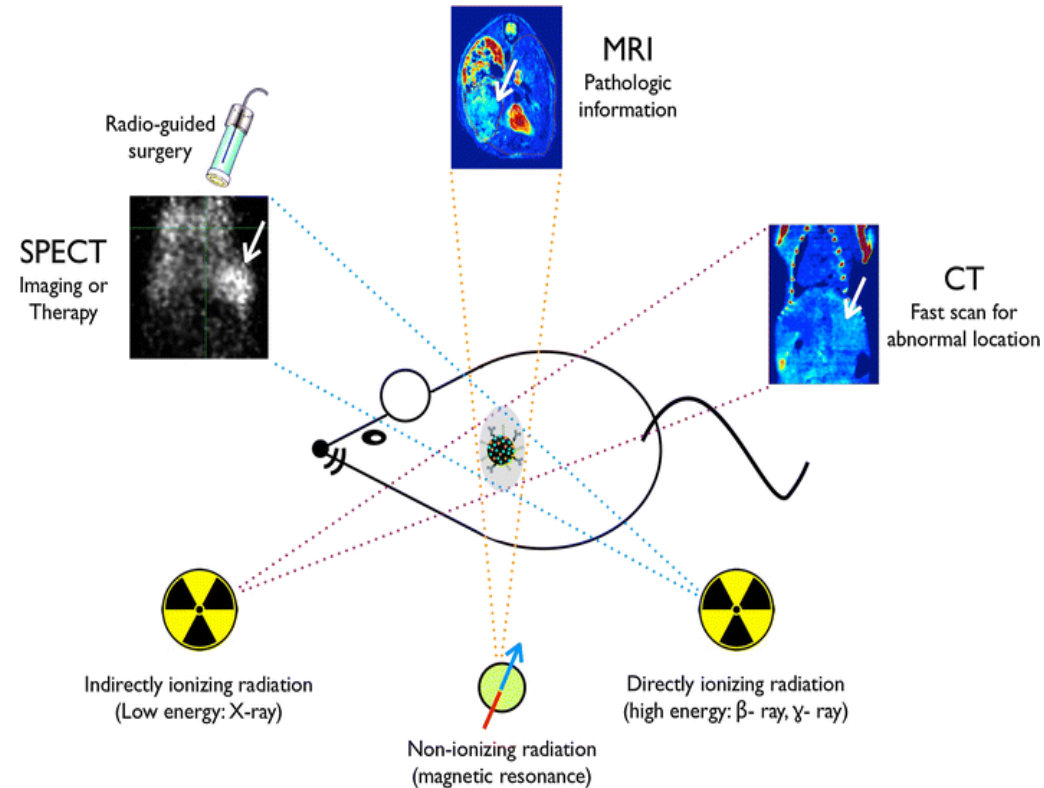


Molecular Screening Applications BIOEMTECH



Conclusions

- Radiolabeling of the nanoparticles may be tricky
 - There are lot of parameters to be taken into consideration.
- Imaging technologies are powerful tools for evaluating the biodistribution of different nanoparticles.
 - They provide unique non invasive tools for repeated studies over time
- It is important to understand the advantages of each imaging technology as well as the properties of different nanoparticles



Our team...



Team includes

- Biomedical Engineers
- Mechanical Engineers
- Physicists
- Biologists
- Radiochemists
- Software Developer
- Software Engineer
- Project Management

Thank you for your attention

